Cross Talk Between the Inflammation and Coagulation Systems

J LYNNE WILLIAMS

ABBREVATIONS: APC = activated protein C; AT = antithrombin; C4bBP = C4b binding protein; DIC = disseminated intravascular coagulation; EC = endothelial cells; EPCR = endothelial cell protein C receptor; GAG = glycosaminoglycans; PAF = platelet activating factor; PAI-1 = plasminogen activator inhibitor-1; PARs= protease activated receptor; PC = protein C; PDGF = platelet derived growth factor; TAFI = thrombin activated fibrinolysis inhibitor; TFPI = tissue factor pathway inhibitor; TM = thrombomodulin; TNF = tumor necrosis factor; ULVWF = ultra-large multimers of von Willebrand factor.

INDEX TERMS: coagulation; cytokines; endothelial cells; hemostasis; inflammation; leukocytes.

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LEARNING OBJECTIVES

- 1. Identify the major components (cellular and inflammatory mediators) of inflammation.
- 2. Describe the functions of the inflammatory response.

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- 3. List the major inflammatory cytokines.
- 4. Identify the significant effects of the inflammatory mediators on the coagulation and fibrinolytic systems.
- 5. Identify the significant effects of the coagulation and fibrinolytic systems on inflammation.

Previously, blood coagulation and inflammation were thought to be completely different physiologic processes. It is now recognized, however, that these two systems are interrelated as part of the host defense mechanism.^{1,2} In invertebrates, the functions of clotting and inflammation are mediated by a single cell system, the hemocyte.3 In vertebrates, clotting and inflammation have diverged into the specialized functions of the platelets, phagocytic cells (neutrophils and macrophages), and several plasma protein systems (procoagulant proteins, complement proteins, proteins of the kinin system).

Several observations suggest an ongoing interaction between these two systems. There is significant structural homology between the proteins of the complement and coagulation cascades. Both cascades utilize serine proteases and are activated through a series of proteolytic cleavage reactions. Animals subjected to experimental sepsis have multi-organ failure associated with activation of blood coagulation (mediated primarily by inflammatory mechanisms). Inhibition of coagulation factor Xa activity inhibits fibrin formation but does not reduce mortality in this model system. However, inhibition of tissue factor/factor VIIa activity or infusion of activated protein C not only reduces fibrin formation, but also significantly reduces mortality.4 Thus, there are interactions between the proteins of these two systems that go beyond simply the activation of fibrin formation.

Both coagulation and inflammation are essential parts of the host defensive response. These pathways have several connecting points through which they interact. Importantly, the interaction between coagulation and inflammation is bidirectional: both inflammation-induced coagulation as well as coagulation-induced inflammation occur.⁵

THE INFLAMMATORY RESPONSE

The inflammatory response refers to the biochemical and cellular processes that occur in vascularized tissue in response

to injury or infection.⁶ It encompasses a complex series of events fundamental to the ability of the human body to protect itself against injurious and infectious agents. The inflammatory response requires the coordinated interaction of 1) local resident cells in the tissues (macrophages, mast cells, fibroblasts); 2) vascular endothelial cells; 3) plasma mediator systems; 4) leukocytes.⁷

The functions of the inflammatory response are: to neutralize or eliminate the offending agent (or to destroy the necrotic tissue); if this cannot be accomplished, the inflammatory response attempts to wall off and confine these agents to limit their effects on the host; to stimulate and enhance the adaptive immune response; and to promote healing.

The classic macroscopic hallmarks of inflammation have been recognized for more than 5000 years and include redness, swelling, heat, and pain. At the microscopic level, these can be explained by the dilation of blood vessels, which increases blood flow into the area, an increase in vascular permeability resulting in the outward leakage of plasma from the vessels into the tissues, and the adherence and emigration of leukocytes through the vascular wall into the surrounding tissues at the site of injury.

The major components of the inflammatory response are listed in Table 1. The process is regulated by a number of inflammatory mediator systems, listed in Table 2. Inflammatory mediators include endogenous (from cells or plasma) as well as exogenous (from bacteria or other pathogens) substances. As can be seen from the table, the list includes a number of components from the coagulation/hemostatic system.

Thus the blood vessels and blood, in addition to being the transport mechanism for the distribution of nutrients and oxygen throughout the body (and the removal of metabolic waste products), are also the essential components of the body's natural immunity (defense) against foreign pathogens and defense against tissue injury. Following injury or infection, an inflammatory response is activated, the integrity of the endothelium is compromised, and phagocytic cells are activated. The endothelium, which is normally anticoagulant and antithrombotic in the resting state, changes to a procoagulant surface, and the balance of the hemostatic system shifts towards hypercoagulability.^{7,8}

INFLAMMATION AND COAGULATION

It has long been recognized that systemic inflammation is associated with hypercoagulability, as evidenced by the

common occurrence of disseminated intravascular coagulation (DIC) in severe sepsis.9 More recently, the molecular basis of this association has been determined. Most of the hypercoagulable effects of inflammation are mediated via the inflammatory cytokines, including IL-1, IL-6 and tumor necrosis factor (TNF).10,11

The hypercoagulability associated with systemic inflammation is due primarily to an increase in procoagulant functions, an inhibition of fibrinolysis, and a downregulation of the three major physiologic anticoagulant systems (activated protein C/APC, antithrombin/AT, tissue factor pathway inhibitor/TFPI, Figure 1):

- 1. Decreased activity of the protein C (PC) anticoagulant system (Figure 2). This is due to decreased endothelial cell expression of thrombomodulin (TM), triggered by IL-6, IL-1 and TNF. 12,13 TM binds thrombin to activate PC to activated protein C (APC). These cytokines also decrease endothelial cell expression of the endothelial cell PC receptor (EPCR), which binds and localizes PC, facilitating its activation by TM/thrombin.¹⁴ In addition, they trigger the hepatic "acute phase response" resulting in increased synthesis and secretion of several acute phase proteins, including C4b binding protein (C4bBP). C4bBP binds plasma protein S, reducing the quantity of "free" protein S. Free protein S is a cofactor for the APC degradation of the procoagulant proteins activated Factor Va and Factor VIIIa.
- 2. Decreased activity of other coagulation inhibitors (Figure 2). Both tissue factor pathway inhibitor (TFPI) and antithrombin (AT) are "negative acute phase response proteins" (meaning their synthesis and secretion is reduced during the acute phase response).⁵ Inflammatory cytokines decrease EC expression of glycosaminoglycans (GAG), the endogenous cofactor for activation of AT activity.¹⁵

Table 1. Components of the inflammatory response

Cells	Plasma protein systems
PMNs (neutrophils) Monocytes/macrophages Eosinophils Basophils/mast cells Platelets	Complement proteins Coagulation proteins Kinin system proteins

- 3. Increased procoagulant activity of the endothelium. Inflammatory cytokines upregulate endothelial expression of tissue factor, the procoagulant protein required for factor VIIa activity,11 and down regulate endothelial expression of heparin
- sulfate/glycosaminoglycans, the cofactors for the anticoagulant activity of antithrombin.15
- 4. Increase in plasma procoagulants. Plasma fibrinogen levels rise, as fibrinogen is another acute phase protein. 16 Inflammatory mediators

induce the endothelial release of ultra-large multimers of von Willebrand factor (ULVWF)—which are hemostatically more effective in inducing platelet activation responses.¹⁷ They also inhibit the cleavage of ULVWF by AD-AMTS13. Inflammatory mediators upregulate TF expression on circulating monocytes, shedding of monocyte-derived microparticles, and initiating systemic coagulation² (Figure 3).

Table 2. Inflammatory mediator systems

Mediator system Major actions

Vasoactive amines: Induction of adhesion, ↑P-, E-selectin Histamine, serotonin Vasodilation; ↑vascular permeability

Vasoactive peptides:

Kinins (bradykinin) Vasodilation, ↑vascular permeability

Endothelial contraction; pain

C3a, C5a †vascular permeability

Chemotaxis; activation of mast cells

Other complement components:

C₃b Opsonization (assist phagocytosis)

C56789 Cytolysis

Coagulation/fibrinolytic:

Fibrinopeptides A, B (FPA,FPB)

Fibrin degradation products (FDP)

Plasmin

†vascular permeability; chemotactic †vascular permeability; chemotactic Generates FDPs; activates C3, factor XIIa, prekallikrein, releases kinins

Thrombin Releases FPA, FPB; †P-, E-selectin; Activate endothelial cells, platelets;

Chemotactic; induce cytokine release

Phospholipids:

Prostaglandins, leukotrienes Vasodilation; ↑vascular permeability;

Chemotaxis

Platelet activating factor (PAF) †vascular permeability; cell activation

Reactive oxygen intermediates:

 O_2 , H_2O_2 , HOTissue damage (cytolysis); activation

of complement; generate chemotactic

lipids

Lysosomal granule contents Tissue damage (proteolysis); matrix

degradation; generate oxidant reactions

Cytokines/chemokines Cell activation; induction of adhesion,

chemotaxis, fever, acute-phase response

system. Inflammatory cytokines stimulate endothelial cells to increase production of plasminogen activator inhibitor-1 (PAI-1).18,5

5. Decreased activity of the fibrinolytic

6. Increased platelet count and platelet reactivity. IL-6 promotes platelet production and is also associated with an increased platelet reactivity (as the newly formed platelets are activated at lower concentrations of agonists).19

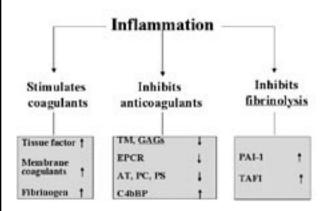
INFLUENCE OF COAGULATION ON INFLAMMATION

Plasma procoagulant enzymes stimulate and augment the inflammatory response. Coagulation inhibitor systems have anti-inflammatory activities in addition to their anti-coagulant functions:

1. Thrombin, factors Xa and VIIa, in addition to their roles in activating coagulation protein zymogens, can interact with specific cell receptors and activate intracellular signaling pathways that mediate inflammatory responses. These enzymes bind protease activated receptors (PARs) on cell membranes and induce a cellular response (Figure 4).^{2,20,21}

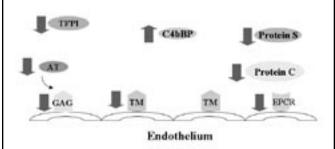
2. Thrombin stimulates endothelial cells (EC), promoting vasoconstriction and increased vascular permeability; promotes adhesion and transendothelial migration of leukocytes; stimulates EC and fibroblasts to produce inflammatory cytokines; activates platelets, inducing the release of platelet-derived inflammatory mediators. 22,23

Figure 1. Impact of inflammatory mediators on coagulation



TM = Thrombomodulin; GAGs = Glycosaminoglycans; EPCR = Endothelial cell protein C receptor; AT = Antithrombin; PC = Protein C; PS = Protein S; C4bBP = Complement component 4b binding protein; PAI-1 = Plasminogen activator inhibitor-1; TAFI = Thrombin activatable fibrinolysis inhibitor

Figure 2. Effects of inflammation on coagulation inhibitor systems



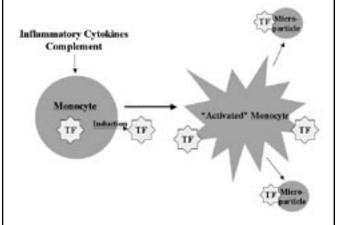
TFPI = Tissue factor pathway inhibitor; AT = Antithrombin; GAG = Glycosaminoglycan; TM = Thrombomodulin; C4bBP = Complement component 4b binding protein; EPCR = Endothelial cell protein C receptor

- 3. Factors VIIa and Xa promote leukocyte adhesion and migration and the synthesis and secretion of inflammatory cytokines.22,24
- 4. Activated protein C (APC) can bind to a PAR, and generate signals that inhibit the inflammatory response (Figure 5).²²
- 5. AT, APC, and TFPI all interact with inflammatory cells and decrease the production of inflammatory mediators, and leukocyte activation in inflammation (Figure 5).^{2,5}
- 6. Thrombin activated fibrinolysis inhibitor (TAFI), in addition to inhibiting fibrinolysis, inactivates proinflammatory mediators including C3a, C5a, and bradykinin (Figure 5). 2,5,25
- 7. Platelets are a rich source of inflammatory mediators; when activated, they secrete a number of factors that promote vascular reactions, leukocyte chemotaxis, endothelial cell activation, and other aspects of the inflammatory response including serotonin, platelet activating factor (PAF), and platelet derived growth factor (PDGF). 1,26

CLINICAL IMPLICATIONS OF "CROSS TALK" BETWEEN INFLAMMATION AND COAGULATION

The processes of coagulation, thrombosis, and inflammation do not happen in isolation. There is a clear interaction

Figure 3. Upregulation of tissue factor on monocytes by inflammatory cytokines



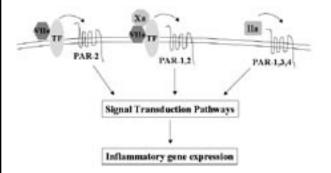
Monocytes are stimulated by cytokines or complement components, triggering tissue factor (TF) synthesis, monocyte shape change and the release of tissue factor-rich microparticles.

between all three, in which thrombosis and coagulation can act as triggers for inflammation, and severe or systemic inflammatory responses can trigger coagulation.

An early outcome of the understanding of the interaction between these processes was the use of the laboratory test, high sensitivity C-reactive protein (hsCRP), as a marker for atherosclerosis and coronary artery disease. 27,28 It is now recognized that an elevated hsCRP may be an early, pre-clinical symptom indication of atherosclerotic inflammatory disease and impending acute thrombotic events. Similarly, studies are underway to evaluate the usefulness of serum IL-6 assays as an indication of the same.

A better understanding of the inflammation/coagulation interface has prompted ongoing research on the interaction of coagulation and inflammation in a variety of disease processes, including inflammatory bowel disease, rheumatoid arthritis, and malignancy. An intriguing study suggesting the importance of the contribution of inflammation to atherothrombosis reported that patients with rheumatoid arthritis

Figure 4. Coagulation proteases and inflammation



Activated coagulation proteases (factor IIa or thrombin, factor VIIa, factor Xa) not only interact with coagulation protein zymogens, but also with specific cell receptors (PARs) to induce signaling pathways that mediate inflammatory responses. PARs are protease activated receptors (of which there are four types, PAR 1 to 4) which, upon cleavage by the coagulation protease, initiates transmembrane signaling, and the induction of genes necessary for the inflammatory response (e.g. synthesis of inflammatory cytokines, cell adhesion molecules, activation of inflammatory responses in macrophages).

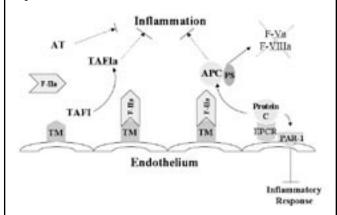
TF = Tissue factor; PAR = Protease activated receptor; VIIa = activated factor VII; Xa = activated Factor X; IIa = thrombin (activated factor II)

exhibited increased risk of myocardial infarction.²⁹ Additional clinical implications of the interaction of these two systems in other disease processes is likely.

An important outcome of the recognition that inflammation and coagulation are intricately related processes is the use of natural anticoagulants as a treatment in acute inflammatory diseases, such as severe sepsis. Activated protein C has been shown to significantly decrease and morbidity and mortality associated with severe sepsis in an animal model, $^{\!4}$ as well as in early human clinical trials. 30 As our understanding of the molecular mechanisms underlying the close relationship between these two systems progresses, it is likely that new potential targets for therapies will be evaluated, capable of modulating excessive activation of both systems simultaneously.

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Figure 5. Anti-inflammatory actions of anticoagulant proteins



The natural anticoagulant proteins also have anti-inflammatory actions. These include antithrombin, activated TAFI, activated Protein C.

TM = thrombomodulin; F-IIa = activated factor II or thrombin; TAFI = Thrombin activatable fibrinolysis inhibitor; AT = Antithrombin; APC = Activated protein C; EPCR = Endothelial cell protein C receptor; PAR-1 = Protease activated receptor-1; F Va = activated factor V; F VIIIa = activated factor VIII

REFERENCES

- 1. Esmon CT, Fukudome K, Mather T, and others. Inflammation, sepsis and coagulation. Haematologica 1999;84:254-9.
- Esmon CT. The interactions between inflammation and coagulation. Br. J Haematol 2005;131:417-30.
- Iwanaga S, Kawabata S, Muta T. New types of clotting factors and defense molecules found in horseshoe crab homolymph: their structures and functions. J Biochem 1998;123:1-15.
- 4. Taylor FBJ, Chang A, Esmon CT, and others. Protein C prevents the coagulopathic and lethal effects of Escherichia coli infusion in the baboon. J Clin Invest 1987;79:918-25.
- Levi M, van der Poll T, Buller HR. Bidirectional relation between inflammation and coagulation. Circulation 2004;109:2698-704.
- Weissman G: Inflammation: Historical perspectives. In: Gallin JJ, Goldstein IM, Snyderman E, editors. Inflammation: Basic Principles and Clinical Correlates, 2nd ed. New York: Raven Press;1992.
- Warren JS, Ward PA. The Inflammatory Response. In: Lichtman MA, Beutler E, Kipps TJ, Seligsohn U, Kaushansky K, Prchal JT, editors. Williams Hematology, 7th ed. New York:McGraw-Hill;2006.
- 8. Levi M, ten Cate H, van der Poll T. Endothelium: interface between coagulation and inflammation. Crit Care Med. 2002;30:S220-4.
- 9. Fourrier F, Chopin C, Goudemand J, and others. Septic shock, multiple organ failure, and disseminated intravascular coagulation. Compared patterns of antithrombin III, protein C and protein S deficiencies. Chest 1992;101:816-23.
- Van Deventer SJ, Buller HR, ten Cate JW, and others Experimental endotoxemia in humans: analysis of cytokine release and coagulation, fibrinolytic and complement pathways Blood. 1990;76:2520-26.
- 11. Levi M, van der Poll T, ten Cate H, and others. The cytokine-mediated imbalance between coagulant and anti-coagulant mechanisms in sepsis and endotoxemia. Eur J Clin Invest 1997;27:3-9.
- 12. Conway EM, Rosenberg RD. Tissue necrosis factor suppresses transcription of the thrombomodulin gene in endothelial cells. Mol and Cell Biol 1988;8:5588-92.
- 13. Nawroth PP, Handley DA, Esmon CT, Stern DM. Interleukin 1 induces endothelial cell procoagulant while suppressing cell-surface anticoagulant activity. Proc Natl Acad Sci USA 1986;83:3460-4.
- Fukudome K, Esmon CT. Identification, cloning and regulation of a novel endothelial cell protein C/activated protein C Receptor. J Biol Chem 1994;269:26486-91.
- Kobayashi M, Shimada K, Ozawa T. Human recombinant interleukin-1 beta and tumor necrosis factor alpha- mediated suppression of heparin-like compounds on cultured porcine aortic endothelial cells. J Cell Physiol 1990;144:383-90.

- Fuller GM, Otto JM, Woloski BM, and others. The effects of hepatocyte-stimulating factor on fibrinogen biosynthesis in hepatocyte monolayers. J Cell Biol 1985;101:1481-6.
- 17. Bernardo A, Ball C, Nolasco L, and others. Effects of inflammatory cytokines on the release and cleavage of the endothelial cell-derived ultra-large von Willebrand factor multimers under flow. Blood 2004;104:100-6.
- Biemond BJ, Levi M, ten Cate H, and others. Plasminogen activator and plasminogen activator inhibitor I release during experimental endotoxaemia in chimpanzees: effect of interventions in the cytokine and coagulation cascades. Clin Sci 1995;88:587-94.
- 19. Burstein SA. Cytokines, platelet production and hemostasis. Platelets 1997;8:93-104.
- Coughlin SR. Thrombin signaling and protease-activated receptors. Nature 2000;407:258-64.
- 21 Camerer E, Huang W, Coughlin WR. Tissue factor- and factor X-dependent activation of protease-activated receptor 2 by factor VIIa. Proc Natl Acad Sci (USA) 2000;97:5255-60.
- 22. Riewald M, Ruf W. Science review: role of coagulation protease cascades in sepsis. Crit Care 2003;7:123-9.
- 23. Strukova SM. Thrombin as a regulator of inflammation and reparative processes in tissues. Biochemistry (Mosc) 2001;66:8-18.
- Schoenmakers SHHF, Reitsma PH, Spek CA. Blood coagulation factors as inflammatory mediators. Blood Cells, Molecules and Diseases 2005;34:30-7.
- 25. Bajzar L, Jain N, Wang P, Walker JB. Thrombin activatable fibrinolysis inhibitor: not just an inhibitor of fibrinolysis. Crit Care Med 2004;32(Suppl 5):S320-4.
- Strukova SM. Role of platelets and serine proteinases in coupling blood coagulation and inflammation. Biochemistry (Mosc) 2004;69:1067-81.
- deBeer FC, Hind CR, Fox KM, and others. Measurement of serum C-reactive protein concentration in myocardial ischaemia and infarction. Br Heart J 1982;47:239-43.
- Danesh J, Wheeler JG, Hirschfield GM, and others. C-reactive protein and other circulating markers of inflammation in the prediction of coronary heart disease. N Engl J Med. 2004;350:1387-97.
- 29. Maradit-Kremers H, Crowson CS, Nicola PJ and others. Increased unrecognized coronary heart disease and sudden deaths in rheumatoid arthritisS: a population-based cohort study. Arthritis and Rheumatism 2005;52:402-11.
- Bernard GR, Vincent JL, Laterre PF, and others. Efficacy and safety of recombinant human activated protein C for severe sepsis. N Engl J Med 2001;344:699-709.

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