

Hemophilia

ABSTRACT:

Hemophilia is a rare, congenital bleeding disorder with an X-linked recessive inheritance pattern. It is characterized by absent, decreased, or dysfunctional coagulation factor FVIII or FIX. Individuals with severe hemophilia bleed into the joints, soft tissue and muscles which can be debilitating. Even though hemophilia is a serious life-long bleeding disorder, understanding the pathogenesis of the disease leads to better patient management and improved patient outcomes.

LEARNING OBJECTIVES:

1. Compare and contrast the differences between hemophilia A and B.
2. Explain the pathogenesis and genetic mechanisms associated with the disease.
3. Describes clinical and laboratory manifestations observed in patients.
4. Lists treatment options for managing patients with hemophilia.

ABBREVIATIONS: AHA - acquired hemophilia A, aPTT - activated partial thromboplastin time, HA - hemophilia A, HB - hemophilia B, HTC - hemophilia treatment center, PT - prothrombin time, TF - tissue factor, VWF - von Willebrand factor.

INDEX TERMS: acquired bleeding disorders, acquired factor VIII deficiency, bleeding diathesis, Christmas disease, bleeding, congenital bleeding disorders, hemophilia, hemophilia A, hemophilia B.

Introduction

Hemophilia is a recessive, X-linked bleeding disorder that results from a decreased or absent level of coagulation factor VIII (FVIII; hemophilia A) or factor IX (FIX; hemophilia B) or to the

inability for either of these coagulation factors to participate in the coagulation cascade. The disorder can be congenital or acquired. The prevalence of congenital hemophilia A (HA) is 1 in 5,000 live male births, and for hemophilia B (HB), 1 in 30,000 live male births.¹ The true prevalence may actually be a little higher due to milder forms of the disorder not being diagnosed in many parts of the world. Initial descriptions of hemophilia date back to as early as the second century in the Talmud stating that male babies should not be circumcised if two previous male siblings died from excessive bleeding as a result of circumcision.² Albucasis later described a family where males died following minor (trivial) injury, and John Conrad Otto is credited with the first modern description of the disorder in 1803.^{2,3} Hemophilia has also been referred to as the “royal disease”, because various members of the royal families were affected by hemophilia. Queen Victoria of England is reported to have been a hemophilia B carrier. She had one son, Leopold, who had hemophilia, and two daughters, Alice and Beatrice, who were carriers and transmitted the disease to the Russian, German, and Spanish royal families (Figure 1).^{1,4} The concept of two different hemophilias was suggested by Pavlovsky when he demonstrated that blood from one hemophilia patient could correct the clotting problem in a second patient with hemophilia, and vice versa.⁵

Hemophilia A and B are clinically indistinguishable from each other, but they differ in the genetic mutation that leads to the bleeding disorder. They also differ in the frequency of inhibitors that develop, quality of life, and patient management. Proper diagnosis can be made by performing coagulation factor assays for FVIII and FIX in the clinical laboratory. In both disorders, the activated partial thromboplastin time (aPTT) is prolonged, while the prothrombin time (PT) is normal, however the activity level of FVIII is decreased/absent in HA and the activity level of FIX is decreased/absent in HB. The hemophilias are further classified as mild

(0.05-0.40 IU/mL, [$>5\%$ to $<40\%$]), moderate (0.01-0.05 IU/mL, [$1-5\%$]), and severe (<0.01 IU/mL, [$<1\%$]), and the bleeding phenotype generally corresponds to the severity of the factor level (Table 1).⁶ Patients with severe hemophilia develop spontaneous and recurrent bleeds without obvious injury or trauma, while patients with moderate and mild hemophilia bleed after injury, trauma, or surgical procedures. The hallmarks of bleeding seen in patients with hemophilia include hemarthrosis (bleeding into the joints) and musculoskeletal bleeding involving the muscles and soft tissue. According to the National Hemophilia Foundation, the percentage breakdown of overall hemophilia population by severity is 60%, 15%, and 25% for severe, moderate and mild deficiencies.⁷

Pathophysiology of Hemophilia

Factors VIII and IX are located in the intrinsic pathway of the coagulation cascade and play a role in the formation of the intrinsic tenase complex upon activation (Figure 2). Factor VIII is known as the antihemophilic factor, and FIX is known as the plasma thromboplastin component or Christmas factor. Immediately following tissue damage, tissue factor (TF) is released into the lumen of the blood vessel and binds to FVII in the circulation to form a TF:VIIa complex. This complex activates FX converting it to FXa and activates FIX to FIXa.⁸ Factor IXa along with FVIIIa converts FX to FXa. Factor VIIIa serves as a cofactor in this reaction increasing the rate of conversion of FX to FXa by FIXa.⁹ Activated FX along with FVa converts prothrombin (FII) to thrombin (FIIa), and thrombin converts fibrinogen to fibrin to form an initial clot. In addition to contributing to fibrin clot formation, thrombin activates FXI to FXIa, FVIII to FVIIIa, and FV to FVa. Factor XIa converts FIX to FIXa. The increased levels of FIXa are now available to interact with the additional FVIIIa leading to continued thrombin generation via the intrinsic pathway. The presence of FVIIIa and FIXa are essential for this continued thrombin generation,

because the extrinsic pathway is downregulated very early during the activation phase of the coagulation cascade. The participation of both extrinsic and intrinsic pathways is needed to form a solid, stable, protective clot. When either FVIII or FIX are absent, severely decreased, or defective, the clot that forms is insufficient to support normal hemostasis.¹⁰ Sixma et al performed histological examinations of clots from patients with hemophilia and demonstrated that the outer region of the clots from hemophiliac patients were stabilized by a fibrin meshwork, whereas the inner portion of the clot showed little or no fibrin formation.¹¹ As a result, hemophilia results from an inability to prolong thrombin generation via the intrinsic pathway due to an absent, decreased, or abnormal production of either FVIII or FIX.

The cell-based model of coagulation adds further insight into the pathogenesis of hemophilia. This model includes platelet and cell surfaces that are essential in *in vivo* coagulation. The cell-based model of coagulation consists of 3 phases to describe coagulation: 1) initiation phase, 2) amplification phase, and 3) propagation phase. During initiation, tissue-factor bearing cells expose TF to FVII in the plasma creating the TF:FVIIa complex that activates FX and FIX. FIXa migrates to the platelet surface while FX remains on the surface of the TF-bearing cell.¹² FXa on the tissue-bearing cell is inhibited by tissue factor pathway inhibitor (TFPI). Amplification occurs on the platelet surface. During amplification, small amounts of thrombin generate activated FV, FVIII, and FXI. The propagation phase follows whereby the intrinsic tenase complex (FVIIIa/FIXa) and the prothrombinase complex (FXa/FVa) form on the platelet surface. FXa on the platelet surface is protected from inhibition by TFPI. A burst of thrombin is generated that converts fibrinogen to fibrin, and activates FXIII which stabilizes the fibrin clot. Formation of both the tenase and prothrombinase complex is critical for prolonged thrombin generation. In patients with hemophilia, an absent/defective FVIII or FIX results in a

nonfunctional intrinsic tenase complex on the platelet surface. In addition, even though FXa is generated in patients with hemophilia, it is not able to move from the tissue-factor bearing cell to the activated platelet surface in order to contribute to thrombin generation.^{12, 13}

Genetics of Hemophilia

Congenital hemophilia A and B are X-linked disorders inherited in a recessive pattern. Sons born to a mother who is a carrier of the mutant gene and an unaffected father have a 50% chance of inheriting the disease, and daughters have a 50% chance of becoming carriers of the mutant gene. All daughters born to an unaffected mother and a hemophiliac father will be carriers of the mutant gene, and none of the sons will be affected (Figure 3a-b).

Additionally, up to 30% of cases may arise *de novo* resulting from a spontaneous mutation in the absence of a family history of hemophilia.¹ There are reports, however, that in up to 80% of the *de novo* mutations, genetic testing of the mothers demonstrated that they were carriers of the mutant gene.¹⁴ The genes encoding FVIII and FIX are located on the tip of the long arm of the X chromosome, Xq28 and Xq27, respectively.^{14,15}

The genes for FVIII and FIX were cloned in 1984 and 1982, respectively.¹⁶ The FVIII gene is 186 kilobases and contains 26 exons that code for a signal peptide and a 2332-amino acid polypeptide. The polypeptide contains 6 domains (A1-A2-B-A3-C1-C2).¹⁷ Factor VIII synthesis occurs in liver sinusoidal endothelial cells and vascular endothelial cells. FVIII circulates as a heterodimer bound to von Willebrand factor (VWF). Upon activation by thrombin, FVIII is released from VWF and interacts with FIXa and FXa on the phospholipid surface of activated platelets.¹⁴ The gene for FIX is 33.5 kilobases and contains 8 exons that code for a 415-amino acid polypeptide.¹⁷ Factor IX is synthesized in liver hepatocytes.¹⁴ Factor

IX is a vitamin K-dependent protein that requires γ -carboxylation before becoming functional. More than 2000 unique molecular defects have been described in the FVIII gene, and about 1,095 genetic variants described in the FIX gene.^{6, 18, 19} The range of mutations for HA and HB include point mutations, insertions, deletions, and inversions. Inversion of intron 22 on the X chromosome is the most common mutation observed in severe HA and accounts for about 45% of cases. The inversion is spontaneous and occurs during meiosis in male germ cells. A similar type of inversion occurs in intron 1 and accounts for 2-5% of cases of severe HA.²⁰ It has been suggested that the reduced frequency of HB compared to HA may be due to the smaller size of the genes involved, 33.5 kb for FIX and 187 kb for FVIII.

Acquired hemophilia

Acquired hemophilia A (AHA) is a rare autoimmune disease and affects approximately 1.5 million individuals annually. It is associated with a high risk for morbidity and mortality.²¹ It is predominantly seen in elderly individuals and equally distributed between males and females. It is also associated with pregnancy and individuals with autoimmune disease. AHA occurs when spontaneous production of neutralizing IgG antibodies target FVIII preventing it from functioning in the intrinsic tenase complex.²² The autoantibodies seen in AHA are different from the alloantibodies that result from exogenous FVIII therapy in patients with congenital HA.²³

Bleeding episodes in patients with AHA often are more severe than in patients with congenital HA, in part due to the kinetics of inactivation of FVIII autoantibodies. Alloantibodies typically inactivate FVIII in a linear fashion (type I, or first-order kinetics) and are concentration- and time-dependent. In addition, alloantibodies completely inactivate FVIII. Autoantibodies, on the other hand, have a rapid inactivation phase followed by a slower equilibrium phase (type II or

second-order kinetics).²³ Autoantibodies incompletely inactivate FVIII. Patients with AHA usually do not have a history of bleeding like patients with congenital hemophilia, therefore early recognition and intervention is essential.

Clinical and Laboratory Manifestations

Clinical symptoms seen in patients with HA or HB are identical with bleeding being the hallmark. The severity and type of bleeding will vary and depends on the severity of the coagulation factor deficiency (Table 1). Bleeding tendencies may also vary with the age of the patient. Neonates with hemophilia may experience intracranial hemorrhage (ICH) at a rate of 40-80 times higher than the normal population.¹⁴ Circumcision may also pose a significant risk of bleeding in cases of severe hemophilia. In children and adults with severe hemophilia, the incidence of musculoskeletal bleeds increases as these individuals are more active and are prone to incidental trauma. These musculoskeletal bleeds contribute to the long-term bleeding complications seen in individuals with severe hemophilia.¹⁴ One way to minimize bleeding in individuals with severe hemophilia is to initiate prophylaxis early on. Prophylaxis involves administration of factor concentrate on a regular basis to maintain adequate factor levels to prevent bleeding. Individuals with moderate to mild hemophilia experience fewer bleeding events and require more significant trauma to initiate bleeding. The amount of bleeding seen in moderate to mild hemophilia correlates with the level of factor activity.

Hemarthrosis is very common and accounts for about 75% of bleeding events in hemophilia.¹⁴

Hemarthrosis tends to repeat in the same joint. The joints most commonly affected are the ankles, knees and elbows. The release of hemoglobin from the erythrocytes ultimately triggers an inflammatory event leading to synovitis. As these events occur on multiple occasions in the

same joint, the synovium undergoes thickening, and the bone and cartilage become destroyed leading to arthropathy.²⁴ Musculoskeletal bleeding and mucosal bleeding (including epistaxis) are commonly seen, as well.²⁵

The development of inhibitors is a serious iatrogenic complication associated with hemophilia treatment. Inhibitors are specific antibodies (IgG) that develop against coagulation FVIII or FIX preparations used in therapy. These antibodies are neutralizing antibodies that inhibit the procoagulant activities of FVIII and FIX. The development of inhibitors is more commonly seen in HA (25 - 30% of patients) compared to HB (3- 5% of patients).²⁴ The lower frequency of inhibitors may in part be due to increased immunogenicity of FVIII compared to FIX.

A prolonged aPTT with a normal PT is the classic finding in the clinical laboratory in patients with hemophilia. The prolongation of the aPTT is directly related to the factor activity levels of FVIII and FIX in the patient. In addition to a prolonged aPTT, laboratories often reflex to a mixing study as part of the algorithm for working up a prolonged aPTT. If an inhibitor has been identified, a laboratory will perform a Bethesda assay to quantitate the antibody titer in the patient sample. A more thorough review of the role of the laboratory in working up hemophilia can be found in the accompanying article Laboratory Monitoring for Hemophilia.

Patient management

Patients who present with bleeding should be treated promptly to avoid severe complications. In addition to being treated for bleeding, these patients should be treated for additional health and psychosocial needs. A hemophilia treatment center (HTC) offers a multidisciplinary approach that takes into consideration various life stages of the patient. The multidisciplinary team includes hematologists, orthopedists, laboratory scientists, nurses, physical therapists, social

workers, and dentists. Treatment and management protocols are individualized based on the patient's age, weight, bleeding pattern, joint health, physical activity, clotting factor levels, and compliance.²⁶ It has been reported that patients utilizing an HTC are 40% less likely to die of a hemophilia-related complication and are less likely to be hospitalized due to bleeding complications.²⁷⁻²⁹ Treatment for bleeding problems may consist of hemostatic support, adjunctive therapies, prophylaxis, and/or inhibitor management. Hemostatic support involves the use of exogenous hemostatic agents to increase the level of deficient FVIII or FIX to an adequate level in the patient to prevent further bleeding. A more in-depth review of the therapeutic agents used to treat patients with hemophilia are found in the accompanying article Past, Present and Future Options in the Treatment of Hemophilia A in this Focus series.

Conclusion

Hemophilia is a very complex bleeding disorder that once was once considered a debilitating disease. During the first half of the 20th century, the life expectancy of patients with hemophilia ranged from about 15 to about 24 years. Today, these patients lead near normal lives due to proper medical care, education about their disease, and the combined efforts of a quality health care team. Great strides have been made in the development of novel hemostatic drugs with extended half-lives that reduce the need for more frequent administration. These drugs have also been optimized for improved safety and efficacy. This has led to measurable reductions in the major complications associated with severe hemophilia. However, the development of inhibitors remains an issue in treating patients with severe hemophilia. The risk of infectious diseases (Human Immunodeficiency, Hepatitis B, and Hepatitis C) transmitted through the use of blood products was very high in the early 80's, but has been eliminated through improved techniques for handling and screening of blood products and the advent of recombinant replacement

products. While these products improve patient compliance and quality of life, they do not provide a “cure.” The only potential cure for hemophilia is gene therapy. However, the challenge remains to develop a safe, viable gene therapy product that is not cost prohibitive. There are additional therapeutic agents on the horizon that down-regulate the naturally occurring inhibitors of coagulation leading to improved hemostasis. These include but are not limited to: 1) molecules that inhibit tissue factor pathway inhibitor; 2) molecules targeting antithrombin; and, 3) a bispecific antibody that targets FIXa and FX mimicking the action of FVIIIa. The progress that has been made in understanding and treating hemophilia is tremendous, but it does not come without cost. The goal will be to continue improvements in safety and efficacy, reduce cost, and increase availability for all individuals worldwide.

Bibliography

1. Mannucci PM, Tuddenham EGD. The Hemophilias – From Royal Genes to Gene Therapy. *N Engl J Med* 2001, 344:1773-1779.
2. Rosner, F. Hemophilia in the Talmud and Rabbinic Writings. *Ann Intern Med* 1969, 70(4):833-837.
3. Franchini, M, Mannucci, PM. Past, Present and Future of Hemophilia: a Narrative Review. *Orphanet Journal of Rare Diseases*, 2012;7:24.
4. Franchini, M, Mannucci, PM. The History of Hemophilia. *Semin Thromb Hemost* 2014;40:571-576.
5. Pavlovsky, A. Contribution to the Pathogenesis of Hemophilia. *Blood* 1947;2(2):181-191.
6. Bhatnagar, N, Hall, GW. Major Bleeding Disorders: Diagnosis, Classification, Management and Recent Developments in Haemophilia. *Arch Dis Child* 2017;0:1-5.

7. Hemophilia A. <https://www.hemophilia.org/Bleeding-Disorders/Types-of-Bleeding-Disorders/Hemophilia-A> , last accessed 6/1/2018.
8. Osterud, B, Papaport SI. Activation of Factor IX by the Reaction Product of Tissue Factor and Factor VII: Additional Pathway for Initiating Blood Coagulation. Proc Natl Acad Sci USA 1977;52:60-4.
9. Kaufman, RJ, Fay, PJ, Popolo, L and Ortel, TL. Factor V and Factor VIII. In: Hemostasis and Thrombosis Basic Principles and Clinical Practice, 6th ed. Lippincott, Wilkins & Wilkins, Philadelphia, PA.
10. Monroe, D, Hoffman, Maureane. What Does It take to Make the Perfect Clot? Arterioscler Thromb Vasc Biol 2006;26:41-48.
11. Sixma, JJ, van den Berg, A. The Hemostatic Plug in Hemophilia A: a Morphological Study of Haemostatic Plug Formation in Bleeding Time Skin Wounds of Patients with Severe Haemophilia A. Br J Haematol 1984;58:741-53.
12. Rominey, G, Glick, M. An Updated Concept of Coagulation with Clinical Implications. JADA 2009;140(5):567-574.
13. Hoffman, M, Monroe III, DM. A Cell-based Model of Hemostasis. Thromb Haemost 2001;85:958-65.
14. Carcao, MD. The Diagnosis and Management of Congenital Hemophilia. Semin Thromb Hemost 2013;38:727-734.
15. Hoyer, L. Hemophilia A. N Engl J Med 1994;330(1):38-47.
16. Giannelli, F, Green, PM. The Molecular Basis of Haemophilia A and B. Baillier's Clinical Haematology 1966;9(2):211-228.

17. Thompson, A. Structure and Function of the Factor VIII Gene and Protein. *Seminars in Thrombosis and Hemostasis*, 2003;29(1):11-22.
18. Factor VIII Gene (F8) Variant Database. <http://www.factorviii-db.org/>, last accessed 6/1/2018.
19. Factor IX Gene (F9) Variant Database. <http://www.factorix.org/>, last accessed 6/1/2018.
20. Swystun, LL, James, PD. Genetic Diagnosis in Hemophilia and von Willebrand Disease. *Blood Reviews* 2017;31:47-56.
21. Kruse-Jarres, R, Kempton, CL, Baudo, F et al. Acquired Hemophilia A: Updated Review of Evidence and Treatment Guidance. *Am J Hematol* 2007;92:695-705.
22. Zdziarska, J, Musial, J. Acquired Hemophilia A: an Underdiagnosed, Severe Bleeding Disorder. *Pol Arch Med Wewn* 2014;124(4):200-206.
23. Kessler, CM, Knobl, P. Acquired Haemophilia: an Overview for Clinical Practice, *European Journal of Haematology* 2015;95 Suppl. 81:36-44.
24. Zimmerman, B, Valentino, LA. Hemophilia: In Review. *Pediatrics In Review* 34(7);289-295.
25. Croteau, SE. Evolving Complexity in Hemophilia Management. *Pediatr Clin N Am* 2018;65:407-25.
26. Dalton, RD. Hemophilia in the Managed Care Setting. *Am J Manag Care*. 2015;21:S123-S130.
27. Soucie, JM, Nuss, R, Evatt, B, Abelhak, et al. Mortality Among Males with Hemophilia: Relations with Source of Medical Care. *Blood* 2000;96:437-442.

28. Soucie, JM, Symons, J, Evatt, B, et al. Home-based Factor Infusion Therapy and Hospitalization for Bleeding Complications Among Males with Hemophilia. *Haemophilia* 2001;7:198-206.
29. Data and Statistics on Hemophilia. <https://www.cdc.gov/ncbddd/hemophilia/data.html>, last accessed 07/06/2018.

Figure 1. The “Royals” Family Tree. Queen Victoria is reported to have been a carrier of hemophilia B. Alice and Beatrice transmitted the disease to the Russian, German, and Spanish royal families.

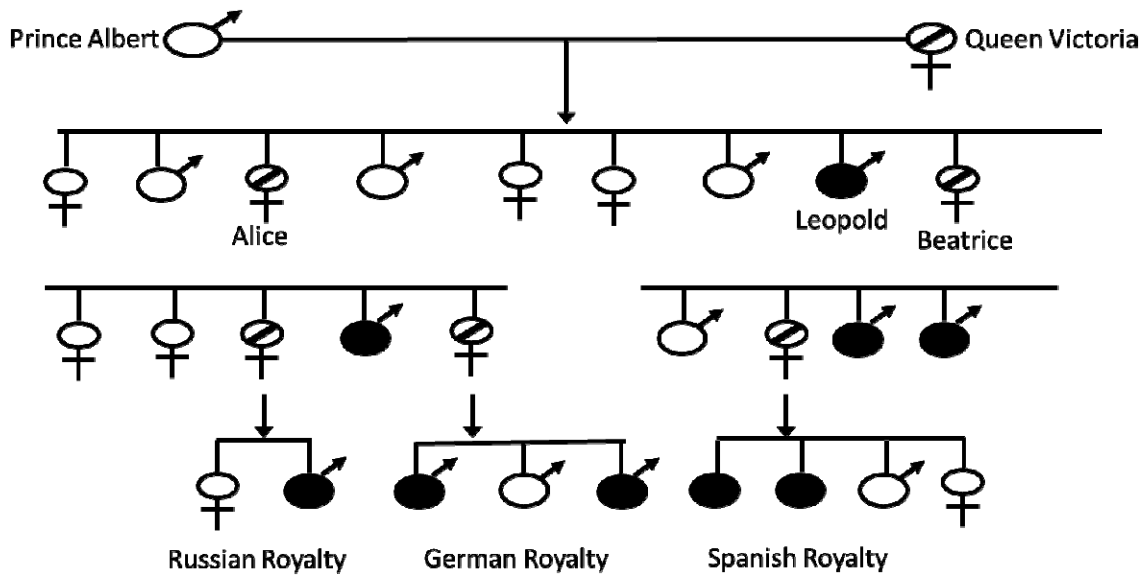


Figure 2. The Coagulation Cascade. FVIII and FVIX are located in the intrinsic pathway.

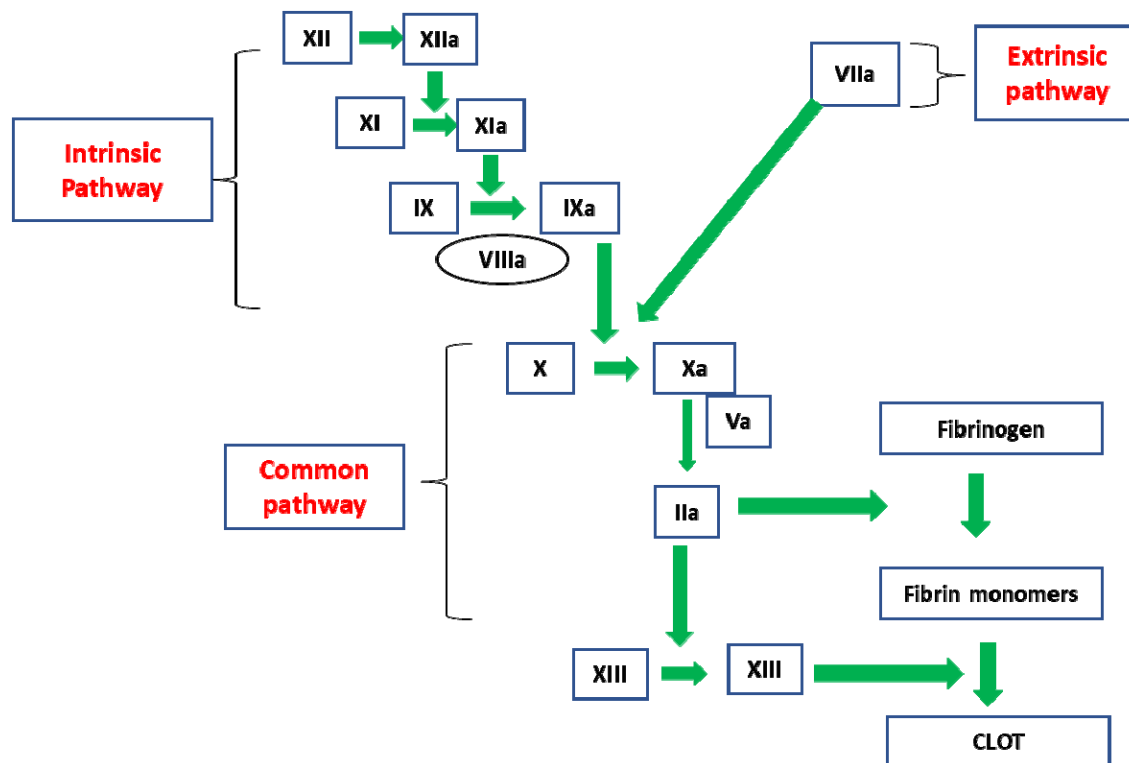


Figure 3a. Inheritance of Hemophilia. Daughters have a 50% chance of inheriting a mutant gene from a carrier mother and unaffected father.

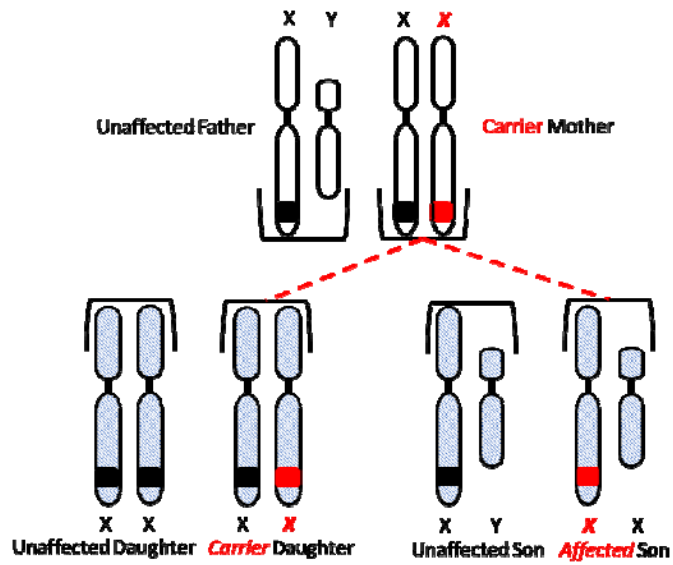


Figure 3b. Inheritance of Hemophilia. All daughters of a affected father and unaffected mother will inherit the mutant gene.

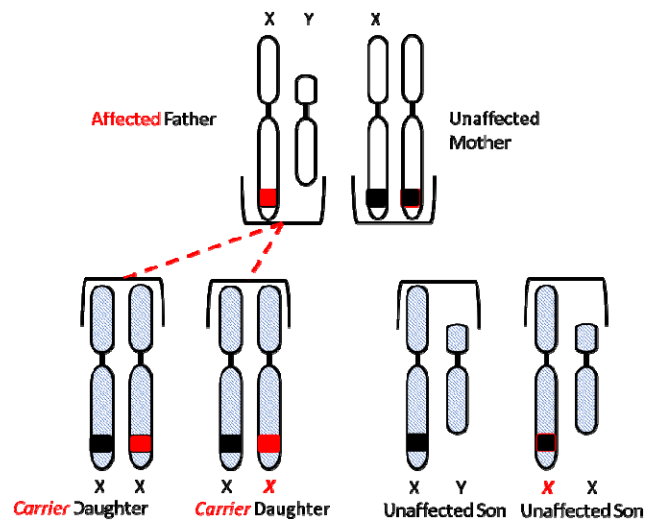


Table 1. Clinical Bleeding Versus Factor Activity.

Severity	Factor Activity Level (%)	Bleeding presentation
Mild	>5	Bleeding after trauma or surgery
Moderate	1-5	Mild trauma or surgery
Severe	<1	Spontaneous